



Effect of *Juglans regia* and *Nasturtium officinalis* on biochemical parameters following toxicity of kidney by CCl₄ in Wistar rats

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ABSTRACT

Background: *Juglans regia* contain compounds such as phenolic acids and flavonoids. Experimental studies demonstrated the *Juglans regia* were used in traditional medicine and has many beneficial effects. Use of *Nasturtium officinalis* has been reported to prevent cellular damage and elevate the level of antioxidant in the body. *Nasturtium officinalis* extract had Strong antioxidant properties and beneficial effects on reducing blood lipids. The purpose of this research is to determine the effect of *Juglans regia* and *Nasturtium officinalis* on biochemical parameters following toxicity of kidney by CCl₄ in Wistar rats.

Materials and Methods: Sixty four Wistar rats were used and divided into 8 groups of eight animals each. Wistar rats were received intraperitoneally 50% CCl₄ in olive oil. *Juglans regia* leaves and *Nasturtium officinalis* were orally administered before or after CCl₄ in treatment groups twice a week for 31 days. After twenty one day, the blood samples were collected of all rats and serum levels of biochemical parameters were measured.

Results: In treatment groups, serum BUN, Bun, Alb, creatinine, Blood Urea, Uric acid levels were significantly decreased in comparison to their positive group. Also at the end of experiment, serum ALP, AST and ALT activity decreased significantly in treatment groups when compared with Positive group.

Conclusion: This study suggests that CCl₄ induced liver and kidney damage in rats can be ameliorated by administration of extract of *J. regia* and *N. officinalis*.

Keywords: liver toxicity, kidney toxicity, Biochemical parameters, *Juglans regia*, *Nasturtium officinalis*

INTRODUCTION

The increase production of Reactive Oxygen Species (ROS) is cause by non-enzymatic reaction in the body and leads to various diseases (1). Different ROS such as hydrogen peroxide, singlet oxygen are unpaired electron and they are extremely reactive molecules (2). They could damage to the biological molecules including DNA, lipids and proteins (1). Furthermore, structural change of proteins leading to the loss of enzyme activity and they have not appropriate function against the free radicals (3). Oxidative stress created when the capacity of the enzymatic system is insufficient to neutralize the free radicals (4). With normal aging the oxidative damage increase to the cell membrane and cause degenerative disease such as cancer, heart and autoimmune disease (3). Recent studies have demonstrated it has been documented the ROS is implicated in many diseases from connective tissue to carcinogenesis (4). It has been shown that carbon tetrachloride (CCl₄) cause triglyceride accumulation in the liver and could create the liver steatosis (5). CCl₄ can disrupted the lipid metabolism and lead the increase the fat droplets in the liver cells (5). Further, it has been documented that CCl₄ leads to production of free radicals that cause different pathological changes in the renal parenchyma such as cell membrane damage, nephrotoxicity and enhance lipid peroxidation (6). Antioxidant has enormous potentials to

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support the different cells from functional damage of toxic radical reaction, drug and oxidative stress (7). They have a key role in prevention of some disease such as cancer, aging, neurological disorder and cataracts (8). Recently, there has been increasing interest in the use of medicinal plants (9). Some medical plants used in many researches to protect the tissue against the oxidative stress and degenerative disease (10).

Traditionally, *Juglans regia* (*J. regia*) was used to treat diabetes, fever, skin diseases and rheumatic pains (11). *J. regia* leaves contained compounds such as phenolic acids and flavonoids, and the most important flavonoids present in the glands of the galactoside and quercetin pentoside derivatives, quercetin arabinoside, quercetin xyloside and quercetin umramnoside (11). *N. officinalis* is full of several protecting vitamins and can be used in the treatment of metabolic and chronic diseases (12). It also prevented cellular damage and increased the level of antioxidants (13). Further, *N. officinalis* is favorable for the evaluation of possible anti-inflammatory effects, antioxidant and antinociceptive properties (13). Since the combination of these two plants has been less studied in blood parameters this study conducted to determine the effects of *Juglans regia* and *Nasturtium officinalis* hydroalcoholic extract on blood parameters following toxicity of liver and kidney by CCl₄ in Wistar rats.

MATERIAL AND METHODS

Sample Preparation and Extraction Procedure

The *J. regia* leaves and aerial parts of *N. officinalis* were obtained during June 2016 from Yasuj region, Yasuj, Iran. The authentication of the *J. regia* and *N. officinalis* plants were confirmed by a taxonomist in the Botany Department of Yasuj University and voucher specimens were 2-12 and 2-13 HMRC respectively. The plants washed thoroughly under running tap water, dried outside in the shade for 5 days and then ground into the fine powder using an electric mixer. The powdered of each plants material (500 g) was soaked in 70% ethanol at room temperature for 24 hours, a procedure repeated twice separately. The mixture was filtered using Whatman No. 1 filter paper. The solvents was removed in a rotary evaporator in vacuum at 57°C and dried for subsequent use. The gained extracts were kept at -20°C in freezer till the time of the experiment.

Animals

Sixty four Wistar rats (6 weeks age, 200-250 g) were procured from the animal house of the Yasuj University of Medical Sciences (YUMS). All experiments were performed according to the local ethical committee in YUMS, for use and care of animals. The animals were maintained under the standard conditions based on ad libitum at room temperature 20±5°C with a regular 12: 12 h light/darkcycle. The animals were divided randomly into eight groups of eight animals. Rats in Group I (negative control) received 0.5 ml/kg body weight of distilled water orally daily and 0.5 ml/kg olive oil by intraperitoneal injection twice a week. Animals in groups 2, 3, 4, 5, 6, 7 and 8 received 1 ml/kg body weight of olive oil and CCl₄ solution twice a week and ratio 1: 1, by intraperitoneal injection. Group 2 (positive control) received 1 ml/kg body weight of olive oil and CCl₄ solution twice a week and ratio 1: 1 by intraperitoneal injection. Group 3 and 4 treated with hydroalcoholic extract of *J. regia* leaves at a dose of 200 and 400 mg/kg gavage daily respectively. Group 5 and 6 treated with hydroalcoholic extract of *N. officinalis* leaves at a dose of 250 and 500 mg/kg gavage daily respectively. Group 7 treated with hydroalcoholic extract of *J. regia* leaves and *N. officinalis* leaves at a dose of 200 and 250 mg/kg gavage daily respectively. Group 7 treated with 50 mg/kg body weight Silymarin standard drug dissolved in normal saline by gavage daily. All groups treated for 31 days.

Serum Biochemistry

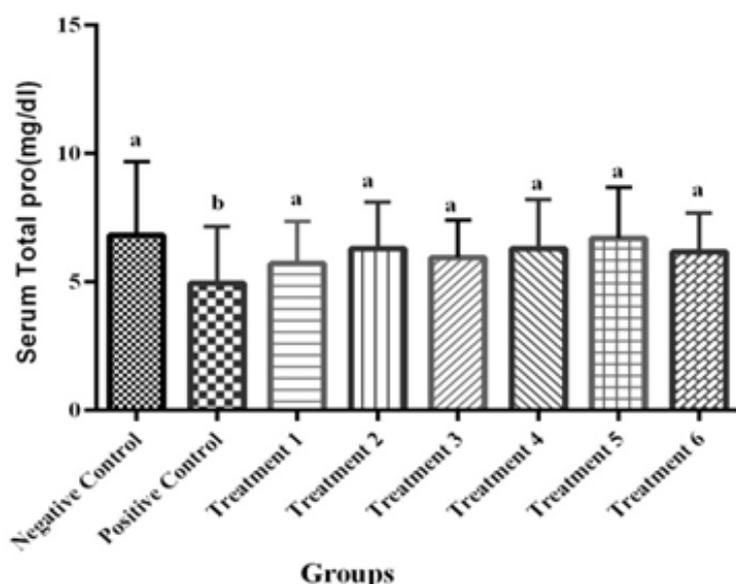
At the end of experimental, the animals were fasted overnight and sacrificed by cervical dislocation after collection of blood samples. Samples were transferred to Hospital's Laboratory for measuring BUN, albumin, Blood Urea, total protein, bilirubin total, bilirubin direct, creatinine, Uric acid and activity levels of AST, ALT and ALP. Blood samples were separated with high speed centrifuge at 3500 rev/minute for 10 minutes and serum was separated by Pasteur pipette for analysis of the biochemical assays.

Statistical Analysis

All data are expressed as mean ± SEM, One-way ANOVA was used for data analysis, followed by the Tukey test for post hoc analysis. A P-value<0.05 was considered to be statistically significant.

Table 1: Effect of *J. regia* leaves and *N. officinalis* on serum activities of ALP, ALT, AST, BUN, Alb, creatinine, blood urea and uric acid on different groups

Groups	ALP(IU/L)	AST(IU/L)	ALT(IU/L)	BUN (mg/dl)	Alb (mg/dl)	Creatinine (mg/dl)	Blood Urea (mg/dl)	Uric acid (mg/dl)
Negative Control	109.50±7.2 ^a	272.66±8.33 ^a	686.00±17.28 ^a	25.34±2.78 ^a	4.10±1.38 ^a	0.63±0.13 ^a	24.61±1.95 ^a	1.06 ± 0.12 ^a
Positive Control	162.50±15.75 ^b	454.66±9.41 ^b	957.16±20.65 ^b	29.66±3.45 ^b	2.01±0.9 ^c	0.93±0.16 ^b	39.96±1.62 ^b	2.11 ± 0.78 ^b
Treatment 1	141.38±9.82 ^a	421.33±6.23 ^b	748.16±16.15 ^a	27.16±2.83 ^a	3.26±1.21 ^a	0.74±0.15 ^a	34.01±2.71 ^b	1.87 ± 0.86 ^b
Treatment 2	121.52±10.66 ^a	344.83±8.49 ^a	729.33±14.39 ^a	26.83±3.37 ^a	3.86±1.47 ^a	0.69±0.12 ^a	26.78±1.45 ^a	1.57 ± 0.62 ^a
Treatment 3	147.16±13.91 ^b	350.16±10.07 ^a	719.16±9.28 ^a	28.02±3.21 ^b	3.11±1.59 ^a	0.82±0.13 ^b	33.20±1.21 ^b	1.91 ± 0.55 ^b
Treatment 4	120.73±12.05 ^a	306.33±8.82 ^a	674.66±12.44 ^a	26.16±2.93 ^a	3.45±1.88 ^a	0.71±0.10 ^a	27.68±1.45 ^a	1.26 ± 0.79 ^a
Treatment 5	111.07±9.27 ^a	275.66±7.95 ^a	759.16±8.75 ^a	23.42±2.05 ^a	3.77±1.25 ^a	0.65±0.14 ^a	25.70±1.90 ^a	1.01 ± 0.33 ^a
Treatment 6	118.33±12.63 ^a	288.50±8.06 ^a	727.33±10.67 ^a	21.00±1.77 ^a	3.46±1.31 ^a	0.73±0.11 ^a	26.05±2.31 ^a	1.39 ± 0.68 ^a

**Figure 1:** The effect of *J. regia* leaves and *N. officinalis* on total protein levels in different groups ($p < 0.05$). The vertical bars are showing mean values of bilirubin total. Lines above the bars indicate standard error (SEM). The word (b) indicates that $P < 0.05$ when comparing positive control group to negative control group. The word (a) indicates that $P < 0.05$ when comparing treated groups to positive control group

RESULTS

Effects of *J. regia* leaves and *N. officinalis* on serum ALT, AST, ALT, Bun, Alb, creatinine, Blood Urea, Uric acid in all groups from various treatment groups are shown in **Table 1**. In Positive group, serum BUN, Bun, creatinine, Blood Urea, Uric acid, ALP, AST and ALT activity were significantly increased in comparison to their Negative group. At the end of experiment, serum ALP, AST and ALT activity decreased significantly in treatment groups of 2, 3, 4 and 5 when compared with Positive group ($p < 0.05$) (**Table 1**).

In addition, **Table 1** shows changes in blood Albumin levels after 31 days. Albumin were decreased significantly in the positive control group (Group II) and brought to the near level as the negative control group in treatment groups ($p < 0.05$). The hydroalcoholic extract of *J. regia* and *N. officinalis* had a significant effect on lowering blood Albumin ($p < 0.05$) (**Table 1**).

The positive control group exhibited, the total protein level showed a significant decrease as compared to the negative group ($p < 0.05$). In addition, the present findings indicate that administration of administrating 200 and 400 mg/kg BW of *J. regia* leaves and 250 and 500 mg/kg bw of *N. officinalis* and silymarin (50 mg/kg bw) lead to significant increase in serum total protein levels in treatment groups as compared with Positive group ($p < 0.05$) (**Figure 1**).

The positive control group exhibited, the bilirubin direct level showed a very highly significant increase as compared to the negative group ($p < 0.05$). The oral administration of *J. regia* and *N. officinalis* produced marked improvement of the bilirubin direct level of the treatment groups as compared with positive group ($p < 0.05$) (**Figure 2**).

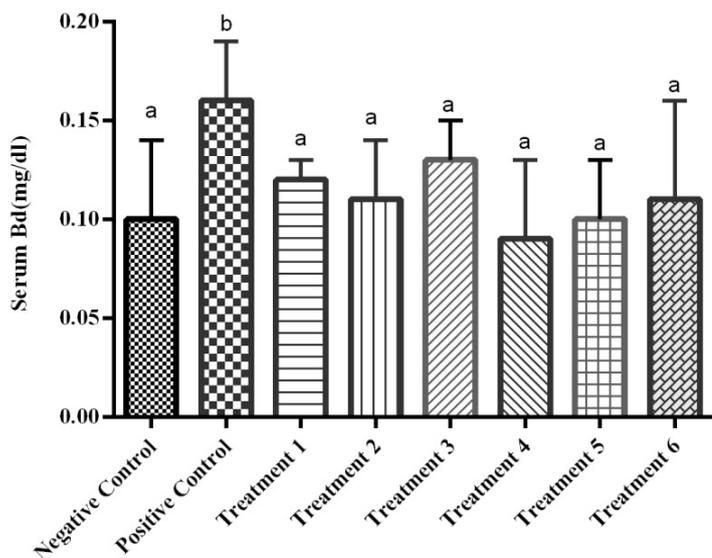


Figure 2: The effect of *J. regia* leaves and *N. officinalis* on bilirubin direct level in different groups ($p < 0.05$). The vertical bars are showing mean values of bilirubin total. Lines above the bars indicate standard error (SEM). The word (b) indicates that $P < 0.05$ when comparing positive control group to negative control group. The word (a) indicates that $P < 0.05$ when comparing treated groups to positive control group

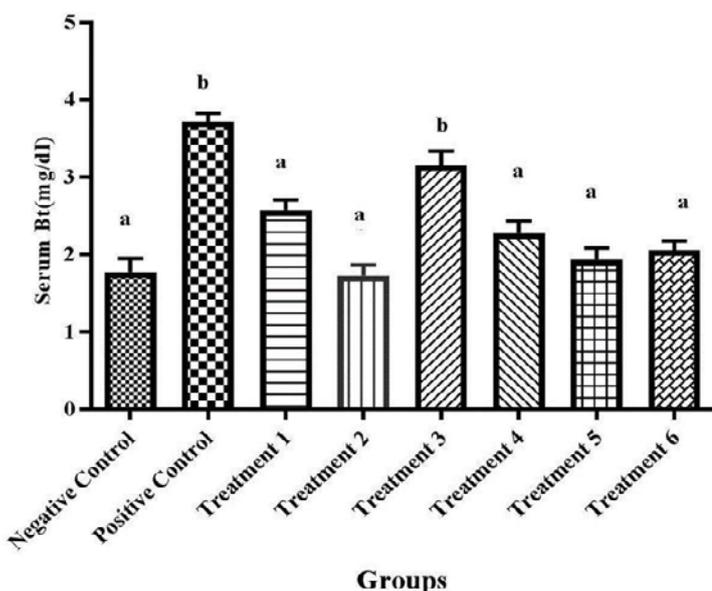


Figure 3: The effect of *J. regia* leaves and *N. officinalis* on bilirubin total levels in different groups ($p < 0.05$). The vertical bars are showing mean values of bilirubin total. Lines above the bars indicate standard error (SEM). The word (b) indicates that $P < 0.05$ when comparing positive control group to negative control group. The word (a) indicates that $P < 0.05$ when comparing treatment groups 1, 2, 4, 5, 6 and 7 to positive control group

Also the positive control group exhibited, the bilirubin total level showed a very highly significant increase as compared to the negative group ($p < 0.05$). The oral administration of *J. regia* and *N. officinalis* produced marked improvement of the bilirubin total level of the treatment groups as compared with positive group ($p < 0.05$) (Figure 3).

DISCUSSION

In the present study the CCl4 treatment caused severe acute liver damage in rats, as evidenced by increased serum AST, ALT, BUN, Bun, Creatinine, Blood Urea, Uric acid and a decreased serum Alb level. Measurement of the activities of

serum marker enzymes, like AST, ALT and ALP, can make assessment of liver function (14). In this study, ALP, AST and ALT activities increased in the Positive group that was treated with CCl₄ alone, indicating damage to the liver tissue by CCl₄ induced hepatotoxicity (p<0.05).

It has been shown that among the different species, the highest activity of catalase and Peroxidase activity was observed in *N. officinalis* (15). The antioxidant potential of plant extracts is related with their phenolic content. Phenolic and flavonoid compounds in plants, have potent antioxidant activity, due to they have a functional group with antioxidant and scavenging activities (16). The hepatotoxic effects of CCl₄ are mostly caused by peroxidation of lipids and the presence of the free radicals (17). In the recent work, the hepatotoxicity was demonstrated by significant increases in the activity of AST, ALT and ALP enzyme markers in CCl₄ treated animals compared to normal group. Determination of ALP activity in plasma is a sign of hepatocyte function (18). In toxic group that treated with CCl₄, the ALP was significantly increased due to damage to liver tissue. However, AST, ALP and ALT activities in the treated with *J. regia* leaves and *N. officinalis* groups were lower than that of the Positive group. This finding suggests that *J. regia* leaves and *N. officinalis* work to protect and repair liver tissue from injury. These results confirm those of previous reports (19, 20). Moreover, our findings suggest that dosages may be an important factor for curative effects of *J. regia* leaves and *N. officinalis*. In another study, oral administration of the hydroalcoholic extract of the *N. officinalis* leaf reduced the serum ALT and AST levels compared with high fat diet groups (20).

An obvious sign of hepatic injury is leakage of cellular enzyme into plasma (21). When the liver cell plasma membrane is damaged, a variety of enzymes normally located in the cytosol are released into blood stream. Their estimation in the serum is a useful quantitative marker for the extent and type of hepatocellular damage (22).

Decrease in serum bilirubin after treatment with the extract in liver damage indicated the effectiveness of the extracts in normal functional status of the liver. This is in agreement with the report by Ahmed *et al.* (23). Hypoalbuminemia and decline in total protein content can be deemed as a useful index of severity of hepatocellular damage. The lowered levels of Albumin and total protein recorded in the serum as well as in the liver of CCl₄-treated rats reveal the severity of hepatopathy (24). From the result of this study, and other findings about antioxidant properties of silymarin, it can be firmly claimed that the antioxidant activity of silymarin leads to improve the change observed in sham rats (25).

Administration of hydroalcoholic extract of *J. regia* leaves increases the HDL and insulin and decreases cholesterol, triglycerides and LDL (16). The *J. regia* extract reduces systolic and diastolic blood pressure. It also increased plasma renin levels and significantly reduced the ratio of aldosterone to rennin (17). In another study, the effect of hydroalcoholic extract of *N. officinalis* leaves on the lipid profile in rats in diabetic rats has been investigated. The results showed that oral administration of hydroalcoholic extract of *N. officinalis* leaves (500 mg / kg) for 10 days reduced serum TC, TG and LDL-C levels respectively, the serum C-HDL level increased (20). Oral administration of the hydroalcoholic extract of the *N. officinalis* leaf reduced the serum ALT and AST levels compared with high fat diet groups. Based on these findings, the *N. officinalis* has a potential for cardiovascular protection and can be used to treat it (18-20).

CONCLUSION

According to the results suggest that the CCl₄ induced liver damage in rats can be ameliorated by administration of hydroalcoholic extract of *J. regia* leaves and *N. officinalis* and improve biochemical factors near to normal.

CONFLICT OF INTEREST

The authors declare no conflict of interests.

REFERENCES

1. Vahidirad M, Arab-Nozari M, Mohammadi H, Zamani E, Shaki F. Protective effect of captopril against diazinon induced nephrotoxicity and neurotoxicity via inhibition of ROS-NO pathway. *Drug Chem Toxicol* 2017;8:1-7. <https://doi.org/10.1080/01480545.2017.1391830>
2. Nazari M, Zarinkamar F, Shafaghat Z. Manganese modulates the physiological and biochemical responses of *Mentha aquatica* L. to ultraviolet radiation. *J Trace Elem Med Biol.* 2018;45:1-10. <https://doi.org/10.1016/j.jtemb.2017.08.015> PMID:29173464
3. Baeri M, Mohammadi-Nejad S, Rahimifard M, Navaei-Nigjeh M, Moeini-Nodeh S, Khorasani R, et al. Molecular and biochemical evidence on the protective role of ellagic acid and silybin against oxidative stress-induced cellular aging. *Mol Cell Biochem* 2017;433:1-13. <https://doi.org/10.1007/s11010-017-3172-0>

4. Okada F. Beyond foreign-body-induced carcinogenesis: impact of reactive oxygen species derived from inflammatory cells in tumorigenic conversion and tumor progression. *Int J Cancer* 2007;121:2364-72. <https://doi.org/10.1002/ijc.23125> PMID:17893867
5. Kato H, Nakazawa Y. Carbon tetrachloride and trichloroethylene toxicities to rat hepatocytes in primary monolayer culture: its relationship to the level of cytochrome P-450. *Toxicol Lett.* 1986;34:55-66. [https://doi.org/10.1016/0378-4274\(86\)90145-1](https://doi.org/10.1016/0378-4274(86)90145-1)
6. Borges Sda S, Korn M, Lima JL. Chromium (III) determination with 1, 5-diphenylcarbazine based on the oxidative effect of chlorine radicals generated from CCl4 sonolysis in aqueous solution. *Anal Sci.* 2002;18:1361-6. <https://doi.org/10.2116/analsci.18.1361> PMID:12502090
7. Pawar R, Mohandass C, Sivaperumal E, Sabu E, Rajasabapathy R, Jagtap T. Epiphytic marine pigmented bacteria: A prospective source of natural antioxidants. *Braz J Microbiol.* 2015;46:29-39. <https://doi.org/10.1590/S1517-838246120130353> PMID:26221086 PMCid:PMC4512047
8. Mukherjee S, Thakur G, Kumar BD, Mitra A, Chakraborty C. Long-term effects of a carbohydrate-rich diet on fasting blood sugar, lipid profile, and serum insulin values in rural Bengalis. *J Diabetes* 2009;1:288-95. <https://doi.org/10.1111/j.1753-0407.2009.00050.x> PMID:20923529
9. Mohammadi J, Naik PR. Antidiabetic effects of *Morus alba* in experimentally induced diabetes in Wistar rat. *Biomedicine* 2008;28:112-116.
10. Yuan LP, Chen FH, Ling L, Bo H, Chen ZW, Li F, et al. Protective effects of total flavonoids of *Bidens bipinnata* L. against carbon tetrachloride-induced liver fibrosis in rats. *J Pharm Pharmacol* 2008;60:1393-402. <https://doi.org/10.1211/jpp.60.10.0016> <https://doi.org/10.1211/jpp/60.10.0016> PMID:18812033
11. Delaviz H, Mohammadi J, Ghalamfarsa Gh, Mohammadi B, Farhadi N. A review study on Phytochemistry and pharmacology applications of *Juglans regia* plant. *Pharmacognosy Reviews* 2017;11:57-64.
12. Mohammadi J, Taheri Motlagh F, Mohammadi N. The effect of hydroalcoholic extract of watercress on parameters of reproductive and sex hormones on the diabetic rats. *J Pharm Sci Res.* 2017;9:1334-38.
13. Shahani S, Behzadfar F, Jahani D, Ghasemi M, Shaki F. Antioxidant and anti-inflammatory effects of *Nasturtium officinale* involved in attenuation of gentamicin-induced nephrotoxicity. *Toxicol. Mech. Methods.* 2017;27:107-114. <https://doi.org/10.1080/15376516.2016.1258748> PMID:27825290
14. Porchezian E, Ansari SH. Hepatoprotective effect of *Abutilon indicum* experimental liver damage in rats. *Phytomed* 2005;12:62-64. <https://doi.org/10.1016/j.phymed.2003.09.009> PMID:15693709
15. Keser G, Saygideger S. Effects of lead on the activities of antioxidant enzymes in watercress, *Nasturtium officinale* R. Br. *Biol Trace Elem Res.* 2010;137:235-43. <https://doi.org/10.1007/s12011-009-8573-9> PMID:19967468
16. Mohammadi J, Chatroz B, Delaviz H. The effect of hydroalcoholic extract of *Capparis spinosa* on quality of sperm and rate of testosterone following induction of diabetes in rats. *Journal of Isfahan Medical School* 2014;31:1-11.
17. Banothu V, Neelagiri C, Adepally U, Lingam J, Bommareddy K. Phytochemical screening and evaluation of in vitro antioxidant and antimicrobial activities of the indigenous medicinal plant *Albizia odoratissima*. *Pharm Biol.* 2017;55:1155-1161. <https://doi.org/10.1080/13880209.2017.1291694> PMID:28219296
18. Johnston DE, Kroening C. Mechanism of early carbon tetrachloride toxicity in cultured rat hepatocytes. *Pharmacol Toxicol.* 1998;83:231-9. <https://doi.org/10.1111/j.1600-0773.1998.tb01475.x>
19. El-Hadary AE, Ramadan Hassanien MF. Hepatoprotective effect of cold-pressed *Syzygium aromaticum* oil against carbon tetrachloride (CCl4)-induced hepatotoxicity in rats. *Pharm Biol.* 2016;54:1364-72. <https://doi.org/10.3109/13880209.2015.1078381> PMID:26440388
20. Lee HS, Li L, Kim HK, Bilehal D, Li W, Lee DS, et al. The protective effects of *Curcuma longa* Linn. extract on carbon tetrachloride-induced hepatotoxicity in rats via up regulation of Nrf2. *J Microbiol Biotechnol.* 2010;20:1331-38. <https://doi.org/10.4014/jmb.1002.03010>
21. Shahrokhi N, Hadad M, Shabani M. Effects of aqueous extract of water cress on glucose and lipids plasma in streptozotocin induced diabetic rats. *Pakistan Journal of Physiology* 2009;5:6-10.
22. Schmidt E, Schmidt FW, Mohr J, Otto P, Vido I, Wrogeman, K, et al. Liver morphology and enzyme release. Further studies in the isolated perfused rat liver. In: Keppler (Ed.) *Pathogenesis and Mechanism of Liver Cell Necrosis.* Medical and Technical Publishing Co. Ltd., Lancaster 1975;147.
23. Ansari RA, Tripathi SC, Patnaik GK and Dhawan BN. Antihepatotoxic properties of picroliv, an active fraction from rhizomes of *Picrorhiza kurroa*. *J Ethnopharmacol.* 1991;34:61-68. [https://doi.org/10.1016/0378-8741\(91\)90189-K](https://doi.org/10.1016/0378-8741(91)90189-K)

24. Ahmed MB, Hasona NAS, Selemain HAH. Protective effects of extract from dates (Phoenix dactylifera) and ascorbic acid on thioacetamide-induced hepatotoxicity in rats. Iranian Journal of Pharmaceutical Research 2008;7:193-201.
25. Aniya Y, Koyama T, Miyagi C, Miyahira M, Inomata C, Kinoshita S, Ichiba T. Free radical scavenging and hepatoprotective actions of the medicinal herb, *Crassocephalum crepidioides* from the Okinawa Islands. Biol Pharm Bull 2005;28:19-23. <https://doi.org/10.1248/bpb.28.19> PMID:15635156
26. Pradeep K, Mohan CVR, Gobianand K, Karthikeyan S. Silymarin modulates the oxidant-antioxidant imbalance during diethylnitrosamine induced oxidative stress in rats. Eur J Pharmacol. 2007;560:110-116. <https://doi.org/10.1016/j.ejphar.2006.12.023> PMID:17300777



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