OXIDATIVE STRESSANDANTIOXIDANT STATUS IN BRONCHOALVEOLAR LAVAGE FLUID, PLASMA AND ERYTHROCYTE OF CRITICALLY MIXED ILL WITH RESPIRATORY FAILURE

Sadık Büyükbaş¹, Kürşat Uzun², Elif Demirkapı¹, Kemal Başaralı¹

Selcuk University, Meram Faculty of Medicine, Departments of Biochemistry¹ and Pulmonary Diseases², Konya, Turkey

Aim: Increased oxidative stress is a significant part of pathogenesis of various lung diseases. In this trial, it is aimed to determine the role of oxidative stress in patients receiving mechanical ventilation for respiratory failure (RF).

Methods: The oxidative stress was evaluated by determining plasma, bronchial fluid and erythrocyte levels of malondialdehyde (MDA), superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), ascorbic acid, alpha-tocopherol, uric acid (UA) and nitric oxide (NO) in 25 critically ill patients with RF. Twenty five patients without RF was evaluated as control group.

Results: Oxidative stress (MDA, NO) levels of plasma, erythrocyte and BAL fluid in mixed critically ill with respiratory failure were higher than control group. Antioxidant levels of plasma, erythrocyte and BAL fluid in mixed critically ill with respiratory failure were lower than control group. In all parameters, there was no statistically difference as exitus and survivors in critically mixed patients.

Conclusion: Oxidative stress was higher in respiratory failure than control group.

Key words: Respiratory failure, oxidative stress, antioxidant status

Eur J Gen Med 2008;5(3):140-146

INTRODUCTION

Lung represents a unique tissue for oxidant stress among most organs because it is directly exposed to higher oxygen tensions (1). In the resting state, the balance between antioxidants and oxidants is sufficient to prevent the disruption of normal physiologic functions; however, either increases in oxidants or decreases in antioxidants can disrupt this balance. The state of imbalance is collectively referred to as oxidative stress and is associated with diverse airway pathologies (2). The major oxidants in airways are reactive oxygen and reactive species (ROS/RNS). nitrogen ROS include superoxide, hydrogen peroxide, and hydroxyl radical. RNS include nitric oxide (NO) (3). Antioxidants are the primary defense against ROS/RNS. The antioxidant effect can be either enzymatic (superoxide dismutase (SOD) and glutathione peroxidase (GSH-Px)) or nonenzymatic (vit C, vit E and uric acid) (4).

Correspondence:Dr. Kursat Uzun Selçuk Üniversitesi Meram Tıp Fakültesi Göğüs Hastalıkları AD. Konya Tel/Fax: 903323237121 E-mail: uzunkur@yahoo.com Oxidative stress during critical illness may be related to activation of phagocytes, production of NO, and release of iron and copper ions and metalloproteins. Critical illnesses, such as sepsis or acute lung injury/ARDS, are characterized by a severe production of ROS and other radical species with consequent oxidative stress (5,6).

The aim of the present study was to investigate the status of oxidative stress and antioxidant in mixed critically ill patients with respiratory failure as assessed by the malondialdehyde (MDA), NO, SOD, GSH-Px, vit C, vit E and uric acid measurements in bronchoalveolar lavage (BAL), erythrocyte and plasma.

MATERIALS AND METHODS

A total 50 subjects were examined, 25 with respiratory failure (average 65.6±12.7 years, 17 males and 8 females) and 25 patients without respiratory failure (average 55.4±9.36 years, 19 males and 6 females). The patients

	Mixed critically ill patients	Control group
n	25	25
Sex (M/F)	17/8	19/6
Age, years	65.6±12.7	55.4±9.36
Length of ICU stay, day	13.35±8.4	-
APACHE II	23.42±5.6	4.7±1.2
PCO ₂ , mmHg	67.41±24.6	43.2±4.1
PO ₂ , mmHg	53.7±12.8	81.5±8.4
HCO ₃ , mmol/L	33.25±15.4	22.1±2.1
pH	7.32±0.15	7.42±0.1
WBC, K/uL	14678.5±7361.8	9237.4±2531.2

Table 1. Characteristics of patients and control group

with respiratory failure including chronic obstructive pulmonary disease (COPD) (n: 15), pneumonia (n: 7) and congestive heart failure (n:3) were divided into two groups as exitus (n:15) and survivor (n: 10) patients. Control group was including patients who performed diagnostic bronchoscopy (lung cancer (n: 18), COPD (n: 7)). Bronchoscopy was performed as described previously (7). Control group is heavy smoker.

BAL; BAL was obtained via endotracheal tube. BAL was performed by Combicath[®] (Plastimed Division, Prodimed, Saint-Leu-La-Foret Cedex, France). About 15 ml of fluid recovered from each subject were centrifuged for 10 min at 300 x g to separate the BAL cells from the acellular BAL fluid.

MDA; Malondialdehyde levels were estimated by the double-heating method of Wasowitz (8). MDA, an endproduct of fatty acid peroxidation, reacts with thiobarbituric acid (TBA) to form a coloured complex. The principle of the method is the spectrophotometric measurement of the colour generated by the reaction of TBA with MDA. The concentration of MDA was calculated by the absorbance coefficient of the MDA–TBA complex (absorbance coefficient $e = 1.56 \times 10^5$ L/mol per cm) and is expressed as µmol / L for plasma and BAL, and nmol/gr Hb for erytrocyte.

Total SOD; Total SOD activity (Cu/Zn and Mn) was determined according to the method of Sun et al (9). Briefly, the principle of the method is based on the inhibition of nitroblue tetrazolium (NBT) reduction by the xanthine/XO system as a superoxide generator. Activity was assessed in the ethanol phase of the supernatant after 1.0 mL ethanol/chloroform mixture (5/3, v/v) was added to the same volume of sample and centrifuged at 4000 g for 30 min at 4 °C. One unit of SOD was defined as the enzyme amount causing 50% inhibition in the rate of NBT reduction. Activity was expressed as U/ml for plasma and BAL, and U/mg Hb for erytrocyte in 560 nm wavelent.

Erythrocyte GSH-Px; Erytrocyte Glutathione peroxidase activity was measured according to the method of Paglia and Valentine (10). The enzymatic reaction in the tube that contained reduced nicotinamide adenine dinucleotide phosphate, reduced glutathione, sodium azide and glutathione reductase was initiated by the addition of hydrogen peroxide (H_2O_2) and the change in absorbance at 340 nm was monitored with a spectrophotometer. Hb determination done spectrophotometric was by cyanmethemoglobin method (Drapkin's solution) and results were given gr/dl . Erytrocyte GSH-Px activity is given in U/gr Hb. All samples were assayed in duplicate.

Plazma E vitamin; The estimated procedure of plasma alpha tocopherol is based upon the original Emmerie-Engel tocopherol assay (11). 0.5 ml of plasma, 0.5 ml of absolue ethanol and 1 ml of n-heptan were pipetted into a glassstoppered 15-ml centrifuge tube. The contents of tube were shaken vigorously by hand for 2 min and centrifuged to completely separate the phases. 0.5 ml of the n-heptan phase (upper layer) was pipetted into another centrifuge tube, and then 0.3 ml α - α ý-dipyridyl and 0.1 ml FeCl₂ were pipetted tube. Produced red

	Exitus	Survivor
n	15	10
Age, years	66.6±13.0	62.4±8.09
Length of ICU stay, day	10.0±3.8	21.7±11.5
Mortality rate, %	53.69±17.2	33.9±19.5
APACHE II	25±5.2	19.5±5.5
PCO ₂ , mmHg	64.24±21.56	75.35±33.5
PO ₂ , mmHg	52.09±14.6	57.7±6.8
HCO ₃ , mmol/L	33.25±15.4	32.1±12.1
рН	7.30±0.15	7.37±0.13
WBC, K/uL	14678.5±7361.8	14567±5531.2

 Table 2. Characteristics of Exitus and survivor patients

color was measured at 510 nm as %T. The results were given in μ g/dl.

Plazma C vitamin; Plasma ascorbate levels were determined photometrically with 2,4-dinitrophenylhydrazine to form the red bis-hydrazone. Ascorbic acid in plasma is oxidized by Cu^{+2} to form dehydroascorbic acid, which reacts with acidic 2,4-dinitrophenylhydrazine to form a red bis-hydrazone, which is measured at 520 nm (12).

Uric acide; Uric acid in the plasma was measured using enzymatic spectrophotometric kit method (GD086500, Globe Diagnostics, Milan, Italy). Uric acid is transformed by uricase into allantoin and hydrogen peroxide which, under the catalytic influence of peroxidase, oxidizes the chromogen (4 aminopherazone/ESPT) to form a purple quinoneimine whose intensity of colour is proportional to the concentration of uric acid in the sample. The results were given in mg/dl.

Plazma NO; The levels of plasma NO were measured using colorimetric kit method (Cat. No. CM780001, Cayman Chemical Company, USA). The best index of total NO production is the sum of both NO2 - and NO3-. The Cayman Chemical Nitrate/Nitrite Assay Kit provides an accurate and convenient method for measurement of total nitrate/nitrite concentration in a simple two-step process. The first step is the conversion of nitrate to nitrite utilizing nitrate reductase. The second step is the addition of the Griess Reagents which convert nitrite into a deep purple azo compound. 3 Photometric measurement of the absorbance due to this azo chromophore accurately determines NO2 – concentration. The levels of plasma NO were given μ M/ L.

RESULTS

Table 1 shows characteristics of patients and control group. Table 2 shows characteristics of exitus and survivor patients. The mean MDA levels of plasma, erythrocyte and BAL fluid in mixed critically ill with respiratory failure were 1.61±0.41 µmol/L, 25.6±8.02 µmol/L and 1.29±0.29 µmol/L, respectively. The mean MDA levels of plasma, erythrocyte and BAL fluid in mixed critically ill with respiratory failure were higher than control group (1.39±0.31 µmol/L, 11.8±3.09 µmol/L and 0.94 ± 0.32 µmol/L, respectively) (p<0.05, p<0.0001, p<0.01). The mean NO levels of plasma and BAL fluid in mixed critically ill with respiratory failure were 55.6±11.7 μ M/L and 44.5±9.4 μ M/L, respectively. The mean NO levels of plasma and BAL fluid in mixed critically ill with respiratory failure were higher than control group (33.95±8.4 μ M/L and 27.16±6.8 μ M/L, respectively) (p<0.0001 and p<0.0001).

Mean SOD levels of plasma, erythrocyte and BAL fluid in mixed critically ill with respiratory failure were 6.99±1.27 U/ml, 3.13±1.33 U/mg Hgb and 6.9±2.29 U/ml,

	Exitus	Survivor	
MDAp, µmol/L	1.62 ± 0.37	1.59±0.39	
NOp, µM/L	53.7±10.9	58.5±9.9	
SODp, U/ml	7.46±0.73	6.28±1.38	
C vitp, mg/dl	0.39 ± 0.16	0.42±0.2	
Evitp, µg/dl	0.58 ± 0.17	0.65±0.19	
UAp, mg/dl	4.6±1.5	5.8±1.1	
MDAe, nmol/gr Hgb	24.2±6.5	27.7±9.0	
SODe, U/mg Hgb	3.39±1.3	2.7±0.4	
GSH-Pxe, U/gr Hgb	88.5±24.7	85.2±9.4	
MDAlv, µmol/L	1.29±0.3	$1.29{\pm}0.2$	
NOlv, µM/L	44.5±8.0	44.2±9.7	
SODlv, U/ml	6.8±1.7	7.1±2.4	

Table 3. The mean levels of oxidants and antioxidants parameters in exitus and survivor patients.

p; plasma, e; erythrocyte, lv; lavage

respectively. The mean SOD levels of plasma, erythrocyte and BAL fluid in mixed critically ill with respiratory failure were lower than control group (7.93±1.0 U/ml, 4.02±1.25 U/mg Hgb and 7.7±1.48 U/ml, respectively) (p<0.05, p<0.05, p<0.05). The mean of uric acid, vitamin C, vitamin E levels of plasma in mixed critically ill with respiratory failure were 4.94±1.52 mg/dl, 0.40±0.19 mg/dl and 0.61±0.18 µg/dl, respectively. The mean of uric acid, vitamin C, vitamin E levels of plasma in mixed critically ill with respiratory failure were lower than control group (7.35±2.03 mg/dl, 0.66±0.27 mg/dl and 1.09±0.31 μ g/dl, respectively) (p<0.0001, p<0.01, p<0.0001).

Mean GSH-Px level of erythrocyte in mixed critically ill with respiratory failure was 87.2 ± 22.5 U/gr Hgb. The mean of GSH-Px level of erythrocyte in mixed critically ill with respiratory failure was lower than control group (111.16±32.7 U/ gr Hgb) (p<0.05). In all parameters, there was no statistically difference as exitus and survivors in critically mixed patients (Table 3).

According to all data, oxidative stress was higher in mixed critically ill with respiratory failure than control group. Antioxidant status was lower in mixed critically ill with respiratory failure than control group.

DISCUSSION

Oxygen-derive free radicals play an important role in the development of disease in critically ill patients. Oxidative stress during critical illness may be related to activation of phagocytes (neutrophils, monocytes, macrophages, eosinophils), production of NO, and release of iron and copper ions and metalloproteins. Critical illnesses, such as sepsis or acute lung injury (ALI)/acute respiratory distress syndrome (ARDS) are characterized by a severe production of ROS and other radical species with consequent oxidative stress (13).

The most frequently used method assess lipid peroxidation is the to measurement of thiobarbituric acidreactive substances (TBARSs) since MDA and other aldehydes react with thiobarbituric acid to give a pink fluorescent colour, which can then be measured. Several studies have confirmed the presence of increased TBARSs in patients with systemic inflammatory response syndrome and multiple organ failure (14,15). Bela et al. (16) reported that critically ill patients irrespective of the disease process indicated significantly very high serum levels of MDA at the time of admission. In an other study, the severity of illness by APACHE III was proportionally related to the degree of oxidative stress (17). In our study, the mean MDA and NO levels as evidences of oxidative stress were higher in patients with respiratory failure (RF) than control group. APACHE II score and mortality rate were 23.4±5.7 and 48.0±19.5% in patients with RF, respectively. There is substantial evidence that experimental reactive oxygen species (ROS) and reactive nitrogen species (RNS) may be involved in pulmonary epithelial injury in a variety of pathologic situations. The induction of immune complex alveolitis in rat lungs results in increased alveolar epithelial permeability, which is associated with the presence of elevated concentrations of NO decomposition products in BAL fluid (18). Sittipunt et al (19) found that NO concentrations were significantly higher than normal in the BAL fluid from patients who were at risk for developing ARDS, as well as in the BAL fluid of those with ARDS. In patients who were at risk for ARDS, the NO concentration in BAL fluid was significantly higher than in healthy subjects (19). Sepsis results in a large increase in the production of nitric oxide and superoxide anions within body (20). Strand et al. (21) found a mean level of 144±39µmol/L in septic patients as compared to 20±3 µmol/L in control subjects. In an other study authors found similar findings (22). In our study, there were a patient with sepsis and two patients with ARDS. NO levels of serum and bronchial lavage fluid were higher in mixed critically ill patients than control group.

ROS are balanced by the activities of enzymes and other molecules called antioxidants, which delay or inhibit oxidation of a substrate. Endogenous antioxidant defences are both nonenzymatic (e.g. uric acid, glutathione, bilirubin. thiols and albumin) and enzymatic (e.g. superoxide dismutase, catalase and the glutathione peroxidase). In the normal subject the endogenous antioxidant defences balance ROS production (23,24).

There is a complex endogenous defense system designed to protect tissues from ROS/RNOS induced cell injury. Special enzymes such as superoxide dismutase, catalase, and glutathione peroxidase, glutathione, and vitamins form a network of functionally overlapping defense mechanisms. In critically ill patients there are reduced stores of antioxidant, reduced plasma or intracellular concentrations of free electron scavengers or cofactors, and decreased activities of enzymatic systems involved in the detoxifications of ROS in critically ill patients (25). The circulating antioxidant levels decrease rapidly after insult, trauma, or surgery and remain below normal levels for several days or even weeks (26).

serum In Bela's study, level of superoxide dismutase was lower in critically ill patients (16). Flaring et al. (27) found that plasma glutathione remains depleted in whole blood in ICU. Selected antioxidants were measured including plasma ascorbate, a major plasma antioxidant, and were significantly decreased in patients with ongoing ARDS when compared to healthy control subjects. Interestingly, the levels of *a*-tocopherol, an additional plasma antioxidant, were unchanged when the two groups were compared (28). Schorah et al (29). reported that lower levels of ascorbic acid in patients of critical care were associated with severity of the illness and were not prevented by parenteral nutrition with ascorbic acid. In our study, the levels of ascorbate and α -tocopherol in plasma were lower in mixed critically ill than control group. Another important antioxidant In our study, the levels of antioxidants (SOD, GSH-Px, Vit C, Vit E, and Uric acid) in plasma, erythrocyte and bronchial lavage were lower in mixed critically ill than control patients. In a study, samples of BAL fluid and epithelial lining fluid from patients with ARDS were analyzed for the presence of GSH, and levels of GSH were found to be decreased when compared to samples from healthy control subjects (30). In another study, levels of catalase were found to increase in patients with sepsis (31). In our study, levels of SOD in BAL fluid were decreased in mixed critically ill with respiratory failure when compared to those of control patients.

In conclusion, oxidative stress is a feature of most respiratory diseases, particularly when inflammation is prominent. Both an increase in ROS/RNS and depletion antioxidants are thought to contribute the pathogenesis of oxidative stress; however it is still unclear which species are the most active in respiratory failure. Decreasing of antioxidant levels (SOD, GSH-Px, Uric acid, Vit C and Vit E) in plasma, erythrocyte and BAL fluid was seen in mixed critically ill with respiratory failure when compared to control group. In parallel higher concentration of plasma, erythrocyte and BAL fluid oxidants (MDA and NO) was seen in mixed critically ill with respiratory failure when compared to control group. The imbalance of antioxidant and oxidants was related with high APACHE scores. Several antioxidants, alone or in combination, have been tested in different small randomized, double blind, placebo-controlled trials. However, antioxidant therapy may be useful in decreasing the mortality of respiratory failure. Clearly, additional studies in this area of research are needed.

REFERENCES

- Kinnula VL, Crapo JD. Superoxide dismutases in the lung and human lung diseases. Am J Respir Crit Care Med 2003;167:1600-1619
- Halliwell B, Gutteridge JMC. Free radicals in biology and medicine, 3rd ed. Oxford: Oxford University Press; 1999
- Bowler RP, Crapo JD. Oxidative stress in airways; is there a role for extracellular superoxide dismutase. Am J Respir Crit Care Med 2002;166:S38-S43
- Rahman I, Biswas SK, Kode A. Oxidant and antioxidant balance in the airways and airway diseases. Eur J Pharma 2006;533:222-239
- Gutteridge JM, Mitchell J. Redox imbalance in the critically ill. Br Med Bull 1999;55:49-75
- Oldham KM, Bowen EP. Oxidative stress in critical care: is antioxidant supplementation beneficial. J Am Diet Assoc 1998;98:1001-1008
- Sokolowsly JW, Burgher LW, Jones FL, Patterson JR, Selecky PA. Guidelines for fiberoptic bronchoscopy in adults. Am Rev Respir Dis 1987;136:1066
- Wasowicz W, Neve S, Peretz A: Optimized steps in fluorometric determination of thiobarbituric acid-reactive substances in serum: Importance of extraction pH and influence of sample preservation and storage. Clin Chem 1993; 39:2522-2526.
- Sun Y, Oberley LW, Ying L:A simple method for clinical assay of superoxide dismutase. Clin Chem 1988; 34:497-500.
- Paglia DE, Valentine WN. Studies on the quantitative and qualitative characterisation of erythrocyte glutathione peroxidase. J Lab & Clin Med 1967; 70:158-169.
- Hashim SA, Schuttringer GR. Rapid determination of tocopherol in marco- and microquantities of plasma. Results obtained in various nutrition and metabolic studies. Am J Clin Nutr. 1966;19(2):137-45.

- Omaye ST, Skala JH, Jacob RA. Plasma ascorbic acid in adult males: effects of depletion and supplementation. Am J Clin Nutr. 1986;44(2):257-64.
- Geoghegan M, McAuley D, Eaton S, Powell-Tuck J. Selenium in critical illness. Curr Opin Crit Care 2006;12:136-141
- 14. Gutteridge JM, Mitchell J. Redox imbalance in critically ill. Br Med Bull 1999;55:49-75
- Motoyama T, Okamoto K, Kukita I et al. Possible role of increased oxidant stress in multiple organ failure after systemic inflammatory response syndrome. Crit Care Med 2003;31:1048-52
- Bela P, Bahl R, Sane AS et al. Oxidative stress status: possible guideline for clinical management of critically ill patients. Panminerva Med 2001;43(1):27-31
- Alonso de Vega JM, Diaz J, Serrano E, Carbonell LF. Plasma redox status relates to severity in critically ill patients. Crit Care Med 2000;28:1812-4
- Mulligan MS, Hevel JM, Marletta MA et al. Tissue injury caused by deposition of immune complex is L-arginine dependent. Proc Natl Acad Sci USA 1991;88:6338-42
- Sittipunt C, Steinberg KP, Ruzinski JT et al. Nitric oxide and nitrotyrosine in the lungs of patients with acute respiratory distress syndrome. Am J Respir Crit Care Med 2001;163:503-10
- Taylor DE, Ghio AJ, Piantadosi CA. Reactive oxygen species produced by liver mitochondria of rats in sepsis. Arch Biochem Biophys 1995; 316:70-6
- Strand OA, Leone A, Giercksky KE et al. Nitric oxide indices in human septic shock. Crit Care Med 2000;28:2779-85
- Dhillon SS, Mahadevan K, Bandi V, Zheng Z, Smith W, Rumbaut RE. Neutrophils, nitric oxide, and microvascular permeability in severe sepsis. Chest 2005;128:1706-12
- Halliwell B, Gutteridge JM. The antioxidants of human extracellular fluids. Arch Biochem Biophys 1990;280:1-8
- Wendel A. Enzymatic basis of detoxification. Vol.1 New York: Academic Press, 1980. pp. 333-53
- Therond P, Bonnefont-Rousselot D, Davit-Spraul A, Conti M, Legrand A. Biomarkers of oxidative stress: an analytical approach. Curr Opin Clin Nutr Metab Care 2000;3:373-84
- Metnitz PGH, Bartens C, Fischer M, Fridrich P, Steltzer H, Druml W. Antioxidant status in patients acute respiratory distress syndrome. Intensive Care Med 1999;25:180-5
- 27. Flaring UB Rooyackers OE, Hebert C, Bratel

T, Hammarqvist F, Wernerman J. Temporal changes in whole-blood and plasma glutathione in ICU patients with multiple organ failure. Intensive Care Med 2005;31(8): 1072-8

- 28. Cros CE, Forte T, Stocker R, et al. Oxidative stress and abnormal cholesterol metabolism in patients with adult respiratory distress syndrome. J Lab Clin Med 1990;115:396-404
- Schorah CJ, Downing C, Pripitsi A, Gallivan L, Al-Hazaa A, Sanderson MJ, Bodenham A. Total vitamin C, ascorbic acid and dehydroascorbic acid concentrations in plasma of critically ill patients. Am J Clin Nutr 1996;63:760-5
- Pacht ER, Timerman AP, Lykens MG, et al. Deficiency of alveolar fluid glutathione in patients with sepsis and the adult respiratory distress syndrome. Chest 1991;100:1397-403
- 31. Leff JA, Parsons PE, Day CE, et al. Increased serum catalase activity in septic patients with the adult respiratory distress syndrome. AM Rev Respir Dis 1992;146:985-9